

SUPPLEMENTARY DATA

Preclinical evaluation of miR-15/107 family members as multifactorial drug targets for Alzheimer's disease

Parsi et al.

This files contains:

- 1) Supplementary Methods**
 - 2) Supplementary Figures and legends**
 - 3) Supplementary References**
 - 4) Supplementary Tables and legends**
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SUPPLEMENTARY METHODS

Patient information

All human and mouse studies were approved by the national ethical committee protocols and in agreement with the Université Laval ethical committee. Brain tissue from patients came from the Douglas Bell Canada Brain Bank, Montreal, Canada, and included non-dementia controls and AD cases, based on neuropathological diagnosis. Patient information is available elsewhere [1, 2]. Blocks of tissue from temporal cortex, prefrontal cortex and hippocampus were dissected and snap frozen in liquid nitrogen until use.

SUPPLEMENTARY FIGURES

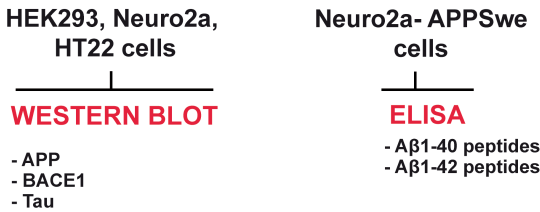
Step 1

Screening and target identification
(all family members)



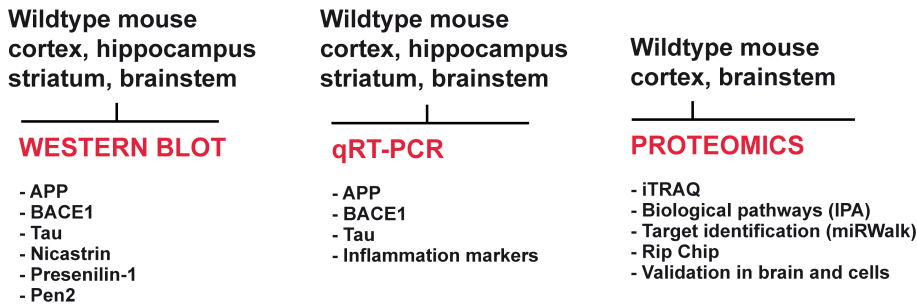
Step 2

Validation and functional studies
(top candidate)

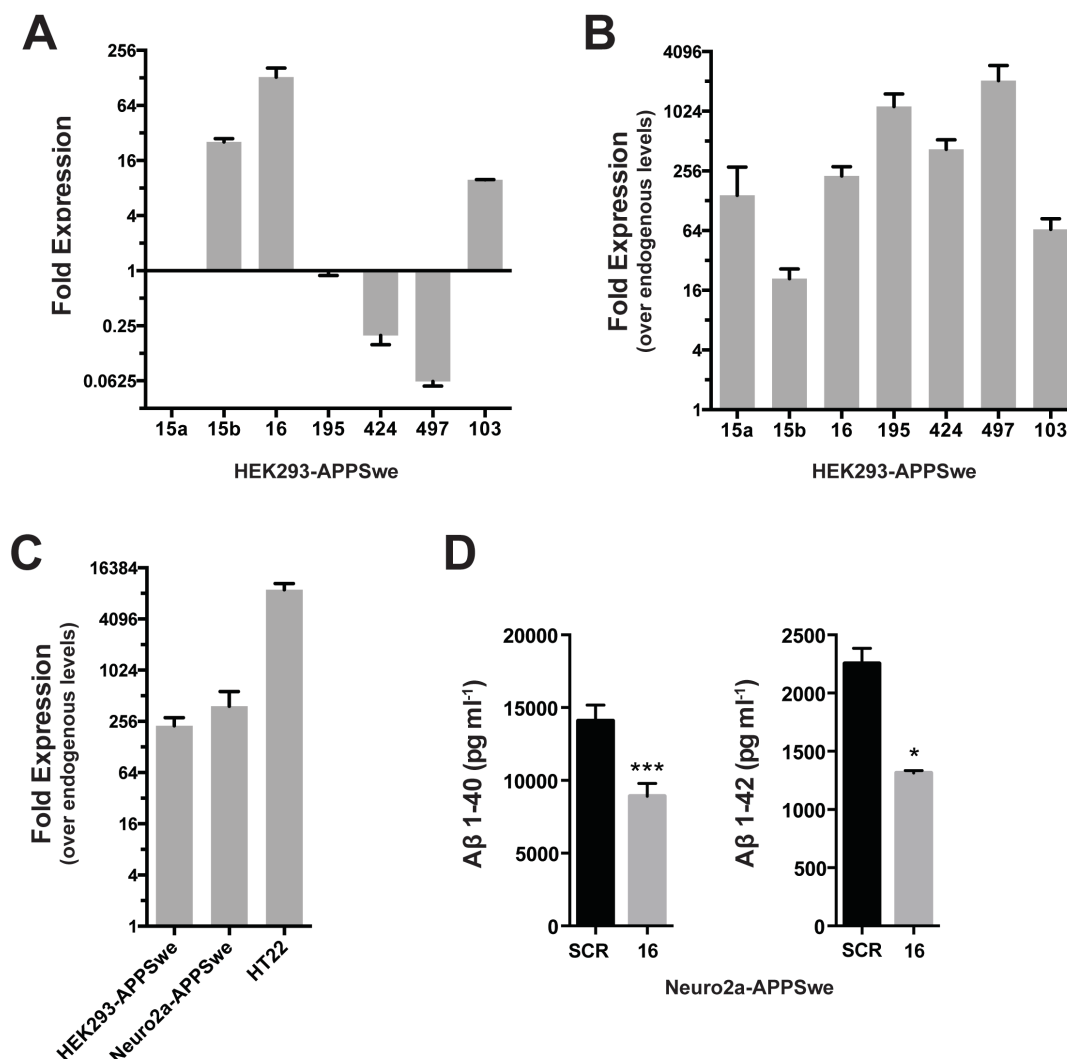


Step 3

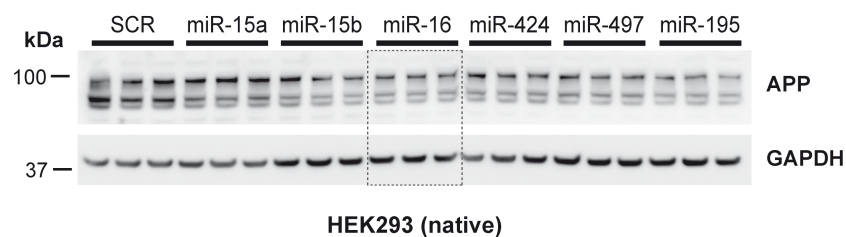
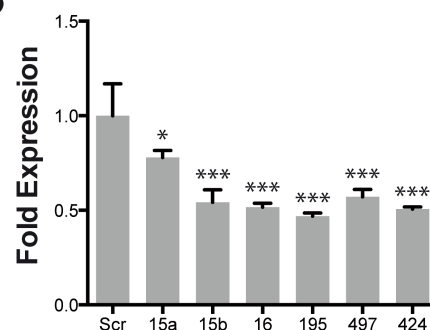
Preclinical proof-of-principle studies
(top candidate)



Supplementary Figure 1. Experimental overview of current study.

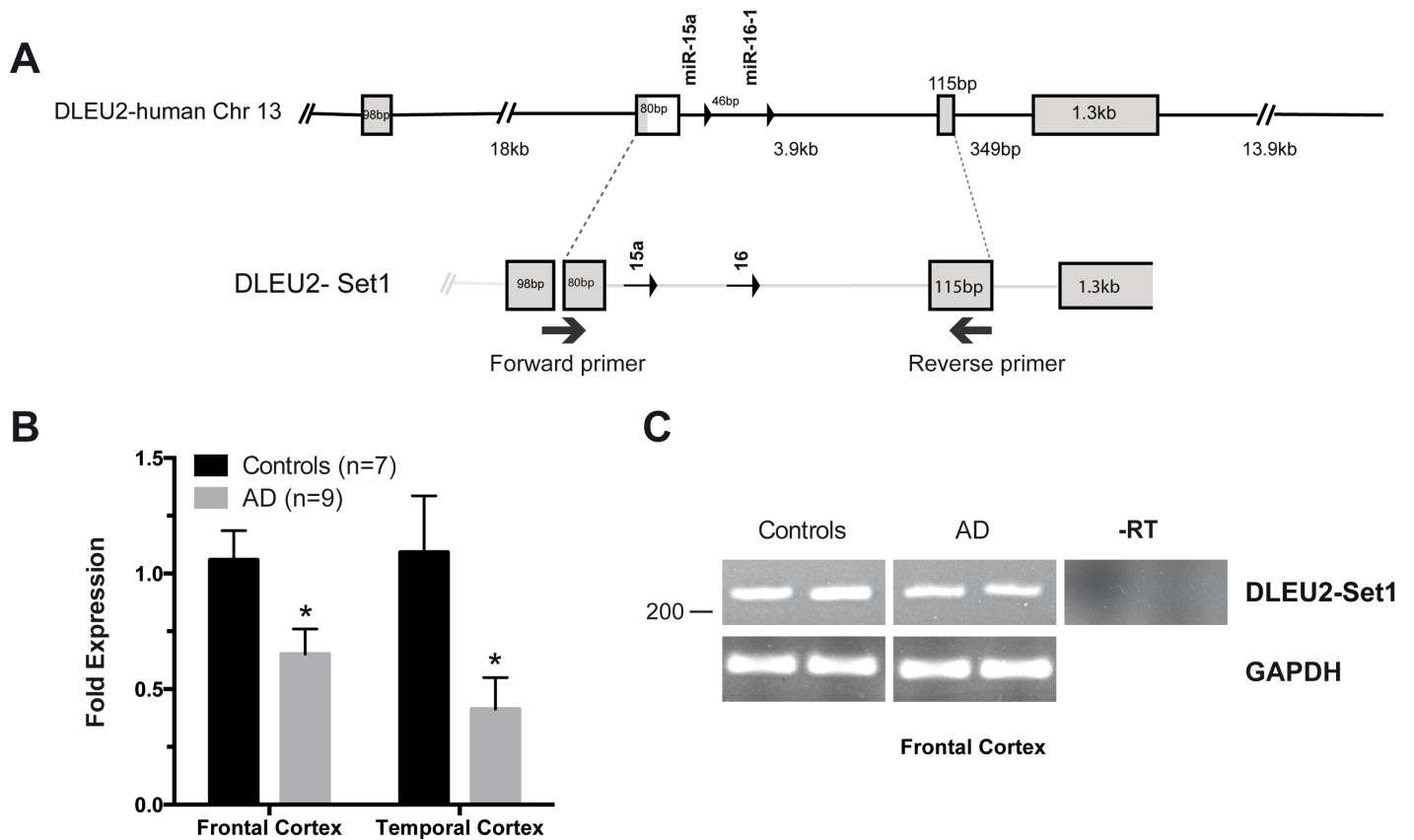


Supplementary Figure 2. (A) Endogenous miR-15a, miR-15b, miR-16, miR-195, miR-424, miR-497 and miR-103 levels were quantified by qRT-PCR in HEK293-APPSwe cells. U48 small nucleolar RNA (RNU48) was used as normalizing control. The relative expression was calculated using the $\Delta\Delta C_t$ method (using miR-15a as 1 fold). (B) qRT-PCR analysis of ectopic miR-16 family members following transfection in HEK293-APPSwe cells. Relative expression is shown (using endogenous miRNAs as 1 fold). RNU48 was used as normalization control. (C) qRT-PCR analysis of transfected miR-16 in various cells lines used in this study. Relative quantifications are shown (using endogenous miR-16 as 1 fold). RNU48 was used as normalization control. (D) ELISA of soluble A β 40 and A β 42 in Neuro2a-APPSwe cells transfected with miR-16 or SCR mimics. Measurements were done 48h post-transfection. Statistical significance was determined by a *Student paired t* test (* = $p < 0.05$, *** = $p < 0.001$). All data are shown as mean \pm SEM from two or more independent experiments in triplicate.

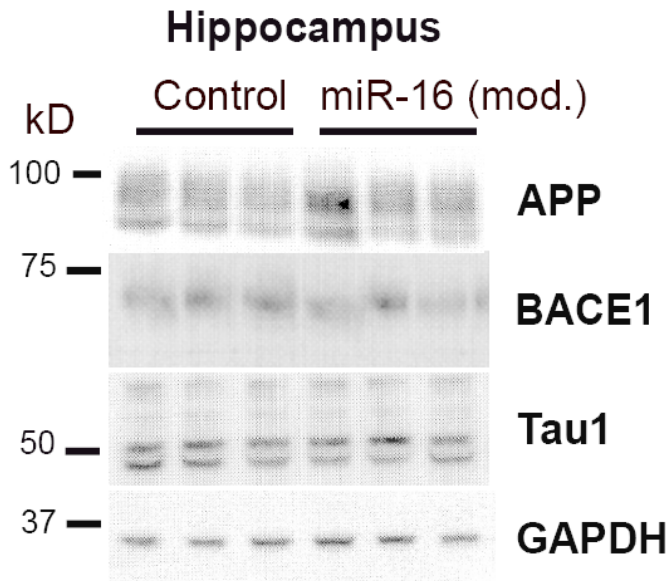
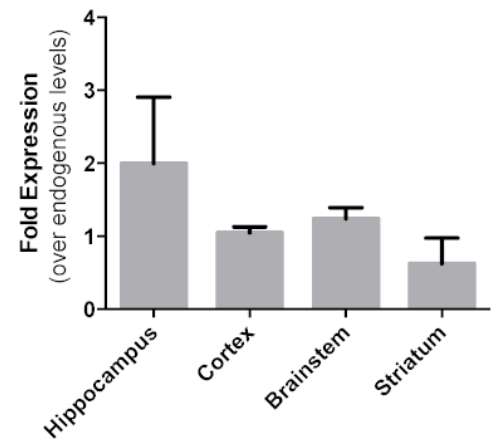
A**B**

Supplementary Figure 3.

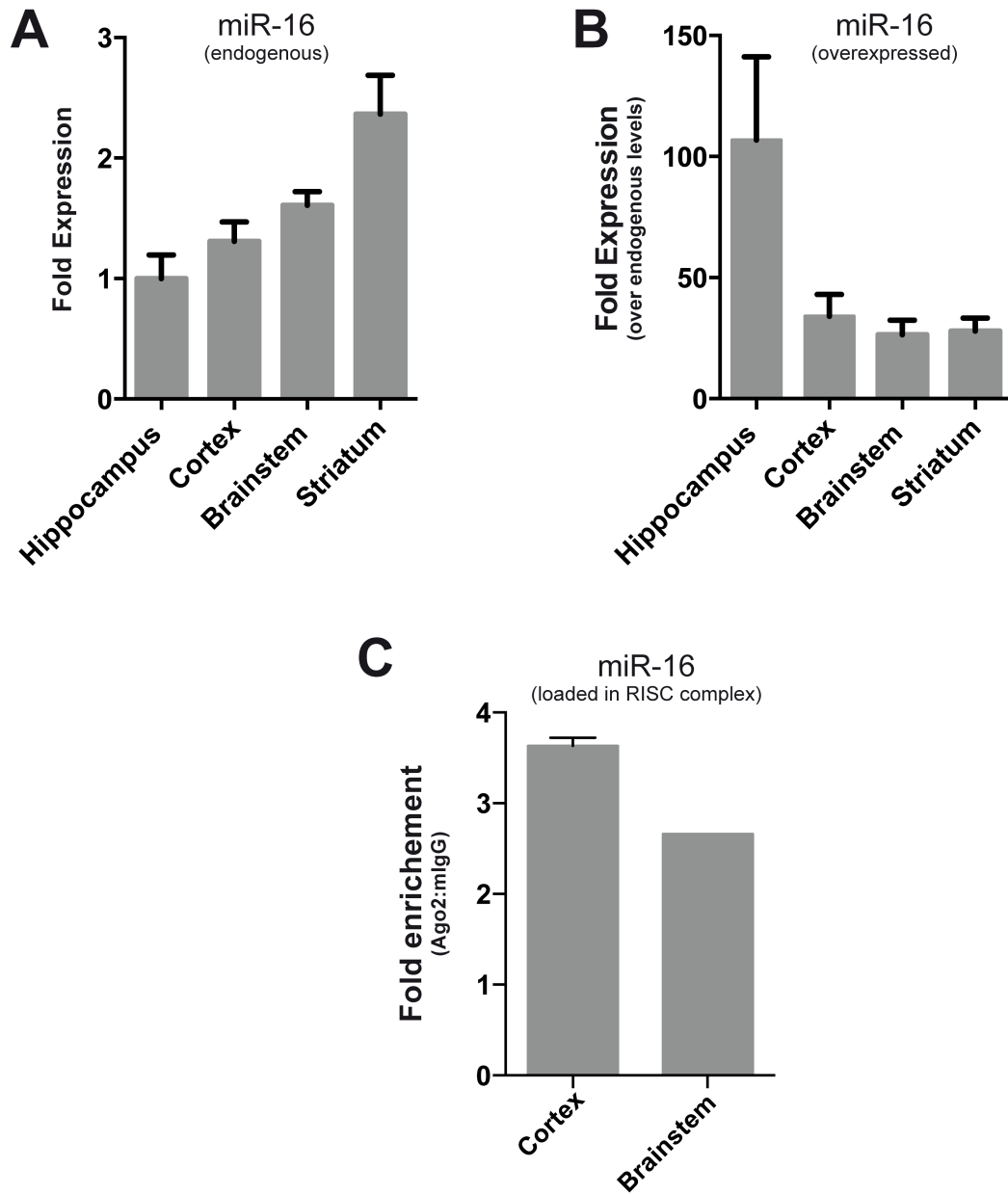
(A, B) Western blot analysis of endogenous full-length APP in native HEK293 cells following mimic overexpression at 50 nM final concentration. Shown here are results at 48h post-transfection. Statistical significance was determined by a *Student paired t* test (* = $p<0.05$, *** = $p<0.001$). Data are shown as mean \pm SEM from two experiments performed in triplicate. Quantifications are shown using Gapdh as normalization control.



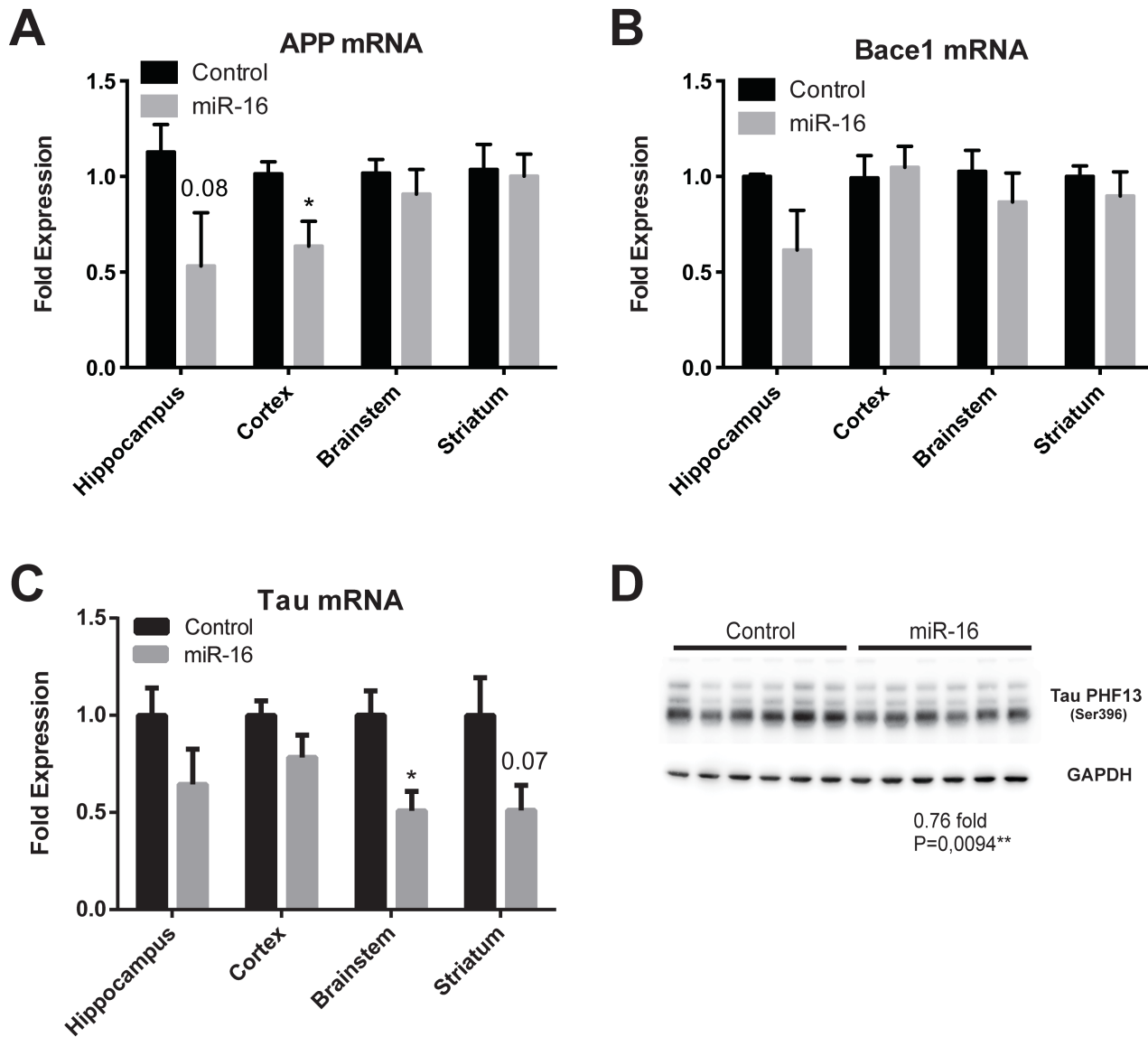
Supplementary Figure 4. Downregulation of the miR-16/15a cluster in AD. **(A)** Schematic representation (not to scale) of the DLEU2 transcript encoding miR-15a and miR-16-1 (upper panel). Close-up of the amplified region (lower panel). Primer sequences are Forward: CTCAGCAATTCTTACCTTTCTTAC; Reverse: TTCCTGGATACTCTCCTGTAGTC. **(B)** qRT-PCR of DLEU2 mRNA from non-demented Controls (N=7) and AD individuals (N=9). Relative expression is shown (using Controls as 1 fold). RNU48 was used as normalization control. All samples were measured in triplicate. Statistical significance was determined by a *Mann-Whitney U* test (* = $p < 0.05$). **(C)** Validation by conventional PCR in frontal cortex tissue. Shown here are two control and two AD individuals. Minus (-) RT was used as PCR negative control.

A**B**

Supplementary Figure 5. Validation of miR-16 mimic specificity *in vivo*. **(A)** Representative western blot analysis of mice treated with chemically-modified miR-16 mimics (miR-16 mod.), harbouring 2O-Me modifications on both sense and antisense stands (50µg/day for 7 days, n=8 mice/group). Shown here is the hippocampus. Similar negative results were obtained in the cortex, brainstem and striatum. Control mice received vehicle alone (0.9% saline) **(B)** qRT-PCR analysis of miR-16 mod.-treated mice. These results indicate that miR-16 expression levels are not significantly increased following treatment (n=8/group), consistent with the notion that modified mimics are not functional. Data are shown as mean± SEM.



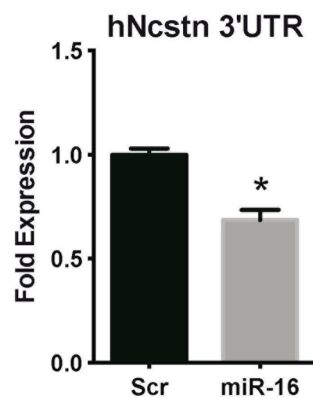
Supplementary Figure 6. (A) Levels of endogenous miR-16 in the different brain regions of control mice (i.e., baseline levels). These experiments were performed from control (saline) treated mice. **(B)** qRT-PCR analysis showing ectopic miR-16 expression and distribution following mimic delivery. These results indicated a strong increase in miR-16 levels (over endogenous levels) in the hippocampus (106 fold), cortex (34 fold), striatum (27 fold), and brainstem (27 fold). **(C)** RIP-Chip was performed on cortex and brainstem of miR-16 mimic-treated mice (n=3/group). We observed a 3.63 and 2.66 fold enrichment in the cortex and brainstem, respectively, compared to controls. Control mouse IgGs served as normalization control. Data are shown as mean \pm SEM.



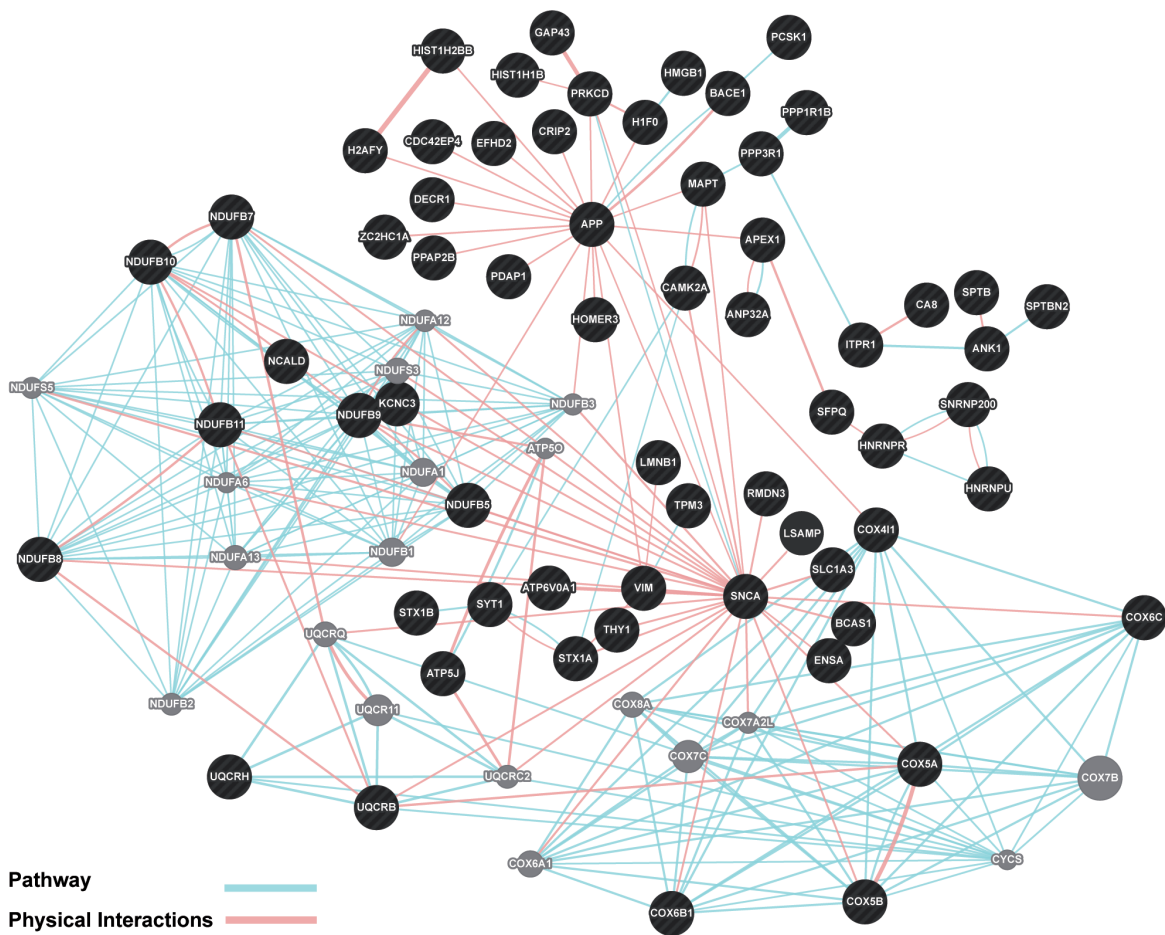
Supplementary Figure 7. Endogenous APP (**A**), BACE1 (**B**), and Tau (**C**) mRNA levels following miR-16 mimic treatment. mRNA levels were measured by qRT-PCR (n=6/group). Statistical significance was assessed by parametric unpaired t test with Welch's correction, $P < 0.05$ considered as statistical significant. GAPDH served as normalization control. Data are shown as mean \pm SEM. (**D**) Representative western blot of endogenous Tau (PHF1 epitope) following miR-16 mimic treatment (N=8/group). Blots were normalized to Gapdh. Statistical significance was determined by a *Student paired t* test.

A

Gene	MicroRNA	DIANAmt	miRanda	miRDB	miRWalk	RNAhybrid	PICTAR5	PITA	RNA22	Targetscan	SUM
NCSTN	hsa-miR-16	1	0	0	0	0	0	0	1	1	3
Ncstn	mmu-miR-16	1	1	0	1	0	1	0	0	1	5

B

Supplementary Figure 8. Nicastrin is directly regulated by miR-16. **(A)** Comparative bioinformatics analysis of putative miR-16 binding sites within the mouse or human Nicastrin 3'UTR. Results were taken from the miRWalk program. **(B)** Luciferase assay on wildtype human Nicastrin 3'UTR co-transfected with 50nM mimics (SCR or miR-16) in HEK293T cells. The cells were lysed 24hrs post-transfection and luciferase signal was measured (n=2 in triplicate). Statistical significance was assessed by one-way ANOVA with Bonferroni post-test (*p < 0.05).



Supplementary Figure 9. Physical interaction networks between putative miR-16 targets *in vivo*. Analysis was performed using the Germania software. Both up- and down-regulated proteins were used in these analyses. Black nodes indicate proteins identified in our proteomics analysis. Grey nodes indicate additional putative binding partners in these pathways based on bioinformatics.

SUPPLEMENTARY REFERENCES

1. Smith, P, Al Hashimi, A, Girard, J, Delay, C, and Hebert, SS (2011). In vivo regulation of amyloid precursor protein neuronal splicing by microRNAs. *Journal of neurochemistry* **116**: 240-247.
2. Hebert, SS, Papadopoulou, AS, Smith, P, Galas, MC, Planel, E, Silahtaroglu, AN, *et al.* (2010). Genetic ablation of Dicer in adult forebrain neurons results in abnormal tau hyperphosphorylation and neurodegeneration. *Human molecular genetics* **19**: 3959-3969.

SUPPLEMENTARY TABLES

Supplementary Table 1. Protein changes in the brainstem and hippocampus of treated mice versus controls (n=4/group). A total of 103 proteins were significantly changed in the brainstem, including 47 upregulated (in red) and 55 downregulated (in green) proteins. A total of 16 proteins were misregulated in the hippocampus, including 5 upregulated (in red) and 11 downregulated (in green) proteins (fold change <0.8 and >1.2, P <0.05). Bioinformatics predictions were done by miRWalk.

Supplementary Table 2. DAVID gene enrichment analysis of top ranked targets identified in brainstem using Homo sapiens background.

Supplementary Table 1

Brainstem										Predicted miR-16 target site	
No Accession	Gene name	Description	Score	95% Coverage	Number of peptides	RATIO mimics vs controls (brainstem)	p-value	3'UTR	Coding region		
Q91XV3	BASP1	BASP1_MOUSE Brain-actin soluble protein 1	82.29	93%	106	0.501	1,65E-07				
Q35262	STX1A	STX1A_MOUSE Syntaxin-1A	24.07	56%	19	0.586	8,92E-06				
P19536	COX5B	COX5B_MOUSE Cytochrome c oxidase subunit 5B, mitochondrial	21.66	53%	16	0.603	0,00121697				
P12787	COKSA	COKSA_MOUSE Cytochrome c oxidase subunit 5A, mitochondrial	39.45	69%	33	0.614	0,000160624				
Q53YK2	Thy1	Thy1_MOUSE Cytoskeleton C90.1	12.01	33%	19	0.618	0,00277828				
Q1MX42	Prkd	Q1MX42_MOUSE Protein kinase C delta type	7.59	9%	5	0.65	0,0326449				
Q1PT73	Syt1	Q1PT73_MOUSE Putative uncharacterized protein	53.45	58%	48	0.653	9,97E-05				
A2AQZ5-4	Skt	A2AQZ5-4 Isoform 4 of Sicke tale protein	11.19	7%	8	0.673	0,00476251				
Q9D6J5	NduB8	NduB8_MOUSE NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 8, mitochondrial	10	38%	9	0.674	0,00526244				
Q4VBC9	Ndufb11	Q4VBC9_MOUSE Ndufb11 protein (Fragment)	10.71	41%	6	0.679	0,0095815				
Q8CPQ1	COW6	COW6_MOUSE Cytochrome c oxidase subunit 6C	17.85	63%	11	0.685	0,000203329				
P56381	Cox6b1	Cox6b1_MOUSE Cytochrome c oxidase subunit 6B1	13.62	70%	20	0.685	0,00071406				
Q9B055	QCR7	QCR7_MOUSE Cytochrome b-c1 complex subunit 7	17.77	57%	15	0.695	0,000153512				
P31650	Sic6a11	56A11_MOUSE Sodium- and chloride-dependent GABA transporter 3	20.84	16%	12	0.698	0,00502416				
Q9JKC6	Cend1	CEND_MOUSE Cell cycle exit and neuronal differentiation protein 1	15.54	57%	19	0.7	0,000423472				
Q60829	Ppp1r1b	PPR1B_MOUSE Protein phosphatase 1 regulatory subunit 1B	18	64%	12	0.702	0,0004223				
P19783	Coxv1	Coxv1_MOUSE Cytochrome c oxidase subunit 4 isoform 1, mitochondrial	14.86	43%	19	0.706	0,0155639				
P06837	Gap43	NEUM_MOUSE Neuromodulin	36.36	71%	32	0.708	5,42E-07				
Q9CQH3	NduB5	NduB5_MOUSE NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 5, mitochondrial	10	21%	6	0.711	0,0077551				
Q62WU2	Tmsb4x	Q62WU2_MOUSE Thymosin, beta 4, X chromosome	12.85	77%	11	0.712	0,0218587				
Q9CR61	NduB7	NduB7_MOUSE NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 7	14.31	58%	8	0.718	0,0129444				
Q8BGT1	Hpc4a1	HPCL4_MOUSE Hippocampin-like protein 4	18.91	81%	23	0.722	0,000985372				
Q9D2C2	CRIP2	CRIP2_MOUSE Cysteine-rich protein 2	10.28	39%	7	0.723	0,0114881				
Q8BLU7	Sic2a3	CRIBL7_MOUSE Putative uncharacterized protein	16.87	13%	11	0.731	0,00105785				
P12798	Camk2a	CKC2A_MOUSE Calcium/calmodulin-dependent protein kinase type II subunit alpha	59.38	50%	75	0.731	0,00178941				
Q9DVM0	PCSK1	PCSK1_MOUSE ProSAs	16.38	47%	12	0.732	0,000728801				
Q91X97	NCALD	NCALD_MOUSE Neurocalin-delta	14.01	74%	25	0.735	0,0215997				
Q91YH8	LPP3	LPP3_MOUSE Lipid phosphatase phospholipase 3	11.31	17%	8	0.74	0,0151303				
Q8QYQ3	BCAS1	BCAS1_MOUSE Breast carcinoma-amplified sequence 1 homolog	30.42	72%	23	0.743	1,08E-05				
Q3YTES	Lsmpp	Q3YTES_MOUSE Limbic system-associated membrane protein	20.73	36%	13	0.749	0,0426762				
P60474	ENSA	ENSA_MOUSE Alpha-enkephalin	13.51	70%	12	0.749	0,00174927				
P99028	QCR6	QCR6_MOUSE Cytochrome b-c1 complex subunit 6, mitochondrial	12.35	60%	8	0.756	0,0278268				
Q3JUX2	Pdp1a	HA2P8_MOUSE 28 kDa heat- and acid-stable phosphoprotein	12.59	34%	9	0.757	0,00129728				
P51830	ADCY9	ADCY9_MOUSE Adenylyl cyclase type 9	6.95	4%	5	0.759	0,0321568				
AS5493	CRYM	CRYM_MOUSE Thiomorpholine-carboxylester dehydrogenase	27.14	51%	23	0.761	0,0168747				
P97450	ATP5J	ATP5J_MOUSE ATP synthase-coupling factor 6, mitochondrial	13.72	56%	12	0.763	0,00373859				
P63321	RALA	RALA_MOUSE Ras-related protein Ral-A	13.51	74%	11	0.765	0,00149347				
Sic17a6	SLC17a6	SLC17a6_MOUSE Vesicular glutamate transporter 2	12.05	17%	8	0.765	0,00936656				
Q9CQZ1	HSBP1	HSBP1_MOUSE Heat shock factor-binding protein 1	8	80%	6	0.765	0,0386879				
Q55042	Snc	SYUA_MOUSE Alpha-synuclein	26.02	91%	31	0.766	0,0013887				
Q63810-2	Ppp3r1	Q63810-2 Isoform 2 of Calcineurin subunit B type 1	26.06	77%	28	0.769	0,00459903				
Q9D883	Chmp4b	CHM4B_MOUSE Charged multivesicular body protein 4b	10.14	29%	9	0.77	0,0219823				
Q95517	SCG2	SCG2_MOUSE Secretogranin-2	46.68	48%	26	0.775	2,77E-07				
NduB10	NduB10	NduB10_MOUSE NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 10	18.01	37%	11	0.777	0,0128557				
Q921G-4	Atg16a1	Q921G-4 Isoform A1-III of V-type proton ATPase 116 kDa subunit A isoform 1	47.89	30%	42	0.783	5,00E-07				
B1A506	Dlgap3	B1A506_MOUSE Disks large-associated protein 3	5.9	5%	5	0.785	0,0338111				
Q8C845	EFhd2	Q8C845_MOUSE EF-hand domain-containing protein D2	24.4	51%	18	0.788	0,000307613				
P13124	KAP3	KAP3_MOUSE cAMP-dependent protein kinase type II-beta regulatory subunit	35.3	52%	18	0.788	0,00215375				
Z2Zc1a	Z2Zc1a	Z2Zc1a_MOUSE Zinc finger C2HC domain-containing protein 1A	12.01	19%	7	0.79	0,0404973				
STX12	STX12	STX12_MOUSE Syntaxin-12	15.15	55%	11	0.79	0,0119414				
STX18	STX18	STX18_MOUSE Syntaxin-18	43.3	56%	38	0.79	1,78E-05				
NduB9	NduB9	NduB9_MOUSE NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 9	16.19	48%	10	0.792	0,0217948				
Q3U2J2	Sic2a1	Q3U2J2_MOUSE Putative uncharacterized protein	10.01	11%	5	0.793	0,0463827				
Q58E70	Tpm3	Q58E70_MOUSE Tpm3 protein	18	49%	22	0.795	0,0242466				
A2A786	HpcA	A2A786_MOUSE Neurexin-specific calcium-binding protein hippocampin (Fragment)	36.76	79%	31	0.795	0,0258137				
Ddc5	Ddc5	Q8BT50_MOUSE DEAD (Asp-Glu-Ala-Asp) box polypeptide 5	45.74	44%	29	1.199	0,0023451				
Ddx39b	Ddx39b	DX39B_MOUSE Spliceosome RNA helicase Ddx39b	27.42	35%	17	1.2	0,00233304				
Sfpq	Sfpq	SFPQ_MOUSE Splicing factor, proline- and glutamine-rich	49.54	42%	35	1.202	0,000109032				
Atg1a1	Atg1a1	ATG1A1_MOUSE Sodium/potassium-transporting ATPase subunit alpha-1	66.56	52%	159	1.203	9,79E-07				
Hnmpr	Hnmpr	Q9CT37_MOUSE Putative uncharacterized protein (Fragment)	12.01	31%	10	1.211	0,0458658				
Hnmpr2	Hnmpr2	HNR12_MOUSE Heterogeneous nuclear ribonucleoprotein U-like protein 2	27.12	26%	17	1.213	0,00892075				
Q92357-5	Ank1	Q92357-5 Isoform 5 of Ankyrin-1	35.26	19%	29	1.223	2,29E-05				
H2afy	H2afy	Q9Q208-2 Isoform 1 of Core histone macro-H2A.1	26.07	38%	15	1.226	0,0200346				
Sorbt2	Sorbt2	SORBT2_MOUSE Sorbin and SH3 domain-containing protein 2	15.78	8%	10	1.227	0,0271742				
Hmgbl	Hmgbl	Q49726_MOUSE High mobility group box 1	15.14	37%	14	1.233	0,0157343				
Phypl	Phypl	PHYPL_MOUSE Phytanoyl-CoA hydroxylase-interacting protein	19.24	42%	15	1.233	0,00591662				
Flna	Flna	FLNA_MOUSE Filamin-A	35.45	12%	23	1.24	0,0234162				
Myh11	Myh11	Q8R384_MOUSE Myh11 protein	32.51	21%	38	1.24	0,00372456				
Coro2b	Coro2b	CORO2B_MOUSE Coronin-28	23.59	31%	13	1.241	0,00286535				
Smpn209	Smpn209	Q9P472_MOUSE Activating signal integrator 1 complex subunit 3-like 1	16.42	7%	13	1.242	0,0415583				
Angp2a	Angp2a	ANG2A_MOUSE Acidic leucine-rich nuclear phosphoprotein 32 family member A	20.06	31%	15	1.242	0,0113832				
Gng13	Gng13	GNG13_MOUSE Guanine nucleotide-binding protein G(i)(G/s)(G/o) subunit gamma-13	6.61	46%	5	1.252	0,0364661				
Decr1	Decr1	DECR1_MOUSE 2,4-dienoyl-CoA reductase, mitochondrial	14.11	24%	7	1.255	0,0164508				
Rmdn3	Rmdn3	RMD3_MOUSE Regulator of microtubule dynamics protein 3	13.61	22%	9	1.258	0,0129001				
Sptb	Sptb	Q3JUG4_MOUSE Putative uncharacterized protein	71.35	29%	57	1.262	6,69E-06				
Arfgef2	Arfgef2	BIG2_MOUSE Brefeldin A-inhibited guanine nucleotide-exchange protein 2	6.74	4%	4	1.264	0,0346823				
G3A3A0	G3A3A0	G3A3A0_MOUSE Heterogeneous nuclear ribonucleoprotein U	36.21	31%	25	1.264	5,20E-05				
Kcnc3	Kcnc3	Q3JZV4_MOUSE Potassium voltage-gated channel subfamily C member 3	11.6	13%	6	1.28	0,0275812				
Lmnbl1	Lmnbl1	LMNB1_MOUSE Lamin-B1	46.05	44%	28	1.285	2,81E-06				
Sptb2	Sptb2	Q68FG2_MOUSE Protein Sptb2	171.89	58%	139	1.286	1,17E-16				
Gpd1	Gpd1	E0C0N5_MOUSE Glycerol-3-phosphate dehydrogenase (NAD+), cytoplasmic	44.7	68%	25	1.29	0,00073079				
Atg2b2	Atg2b2	Q3JUH0_MOUSE Putative uncharacterized protein	80.6	39%	64	1.292	5,28E-07				
Fam88b	Fam88b	FAM88B_MOUSE Protein FAM88B	9.85	17%	5	1.297	0,0259779				
H10	H10	Q3U4Y0_MOUSE Putative uncharacterized protein	12	27%	9	1.297	0,041291				
Vim	Vim	VIME_MOUSE Vimentin	68.44	75%	60	1.316	5,50E-06				
Ehd1	Ehd1	EHD1_MOUSE EH domain-containing protein 1	33.52	62%	29	1.333	0,006632995				
Apgk	Apgk	Q8C413_MOUSE Diacylglycerol kinase gamma	18.4	19%	10	1.347	0,00102144				
Apekl	Apekl	APKL_MOUSE DNA (apurinic or apyrimidinic site) lyase unknown [Mus musculus] [10090]	16.05	38%	10	1.466	0,0323024				
Pvalb	Pvalb	PVALB_MOUSE Parvalbumin alpha	20	77%	12	1.518	0,000198945				
Sic1a3	Sic1a3	Q9NGM1_MOUSE Glutamate/aspartate transporter (Fragment)	21.1	22%	36	1.532	0,0213031				
Cdc42ep4	Cdc42ep4	BORG4_MOUSE Cdc42 effector protein 4	10	16%	5	1.549	0,0279819				
Hist1h3b	Hist1h3b	H15_MOUSE Histone H1.5	8.21	19%	5	1.714	0,02542				
Hist2b2	Hist2b2	H2B18_MOUSE Histone H2B type 1-B	16.08	43%	34	1.722	0,0243785				
Acap2	Acap2	Q6ZQK5-2 Isoform 2 of Arf-GAP with coiled-coil, ANK repeat and PH domain-containing protein 2	6.39	7%	3	1.731	0,0222356				
Q8BQZ8	Q8BQZ8	Q8BQZ8_MOUSE Putative uncharacterized protein	4.96	11%	3	1.751	0,0211423				
Tagln	Tagln	TAGL_MOUSE Transgelin	18.77	52%	11	1.753	0,000255131				
Calb1	Calb1	CALB1_MOUSE Calbindin	25.76	67%	22	1.802	2,99E-08				
Itrp1	Itrp1	P11881-8 Isoform 8 of Inositol 1,4,5-trisphosphate receptor type 1	71.39	19%	48	2.308	6,43E-19				
Homcr3	Homcr3	Q9JP6-2 Isoform 2 of Homer protein homolog 3	8.94	22%	6	3.396	0,000109871				
Sic1a6	Sic1a6	Q3TXM3_MOUSE Putative uncharacterized protein	8	17%	10	3.784	0,000775311				
Ca8	Ca8	CAH8_MOUSE Carbonic anhydrase-related protein	12.02	33%	6	4.709	6,54E-05				

Supplementary Table 2

Background: Home sapiens						
Category	Term	Count	%	P-Value	Benjamini	
KEGG_PATHWAY	Alzheimer's disease	11	15.1	5.1E-10	3.0E-8	
	Parkinson's disease	12	12.9	6.3E-9	1.0E-7	
	Oxidative phosphorylation	12	12.9	7.5E-9	1.5E-7	
	Huntington's disease	12	12.9	1.2E-7	3.3E-6	
	Cardiac muscle contraction	7	7.5	4.8E-5	5.6E-4	
	SNARE interactions in vesicular transport	3	3.2	4.3E-3	3.6E-1	
	SHARK signaling pathway	4	4.3	5.2E-3	3.6E-1	
	Calcium signaling pathway	5	5.4	6.6E-3	3.6E-1	
	Oocyte meiosis	4	4.3	6.9E-3	3.7E-1	
	Vascular smooth muscle contraction	4	4.3	7.2E-3	3.6E-1	

Category	Term	Count	%	P-Value	Benjamini	
GOTERM_BP_FAT	oxidative phosphorylation	8	8.6	1.7E-6	1.7E-3	
	respiratory electron transport chain	7	7.5	2.6E-6	9.8E-4	
	generation of precursor metabolites and energy	12	12.9	3.2E-6	7.0E-4	
	electron transport chain	8	8.6	4.8E-6	1.2E-3	
	mitochondrial ATP synthesis coupled electron transport	6	6.5	1.9E-6	3.7E-3	
	ATP synthesis coupled electron transport	6	6.5	1.9E-6	3.7E-3	
	cellular respiration	7	7.5	3.2E-6	3.7E-3	
	mitochondrial electron transport, NADH to ubiquinone	5	5.4	1.1E-6	1.5E-3	
	neurotransmitter transport	6	6.5	1.3E-6	1.5E-3	
	regulation of cellular component biogenesis	7	7.5	1.9E-6	2.0E-3	
	energy derivation by oxidation of organic compounds	7	7.5	2.1E-6	2.0E-3	
	regulation of synaptic transmission	6	6.5	1.2E-6	1.6E-3	
	oxidation-reduction	12	12.9	1.2E-6	9.8E-3	
	negative regulation of cellular component organization	6	6.5	1.5E-6	1.2E-3	
	regulation of transmission of nerve impulse	6	6.5	1.8E-6	1.2E-3	
	regulation of protein complex assembly	5	5.4	2.0E-6	1.2E-3	
	regulation of neurological system process	6	6.5	2.1E-6	1.2E-3	
	regulation of system process	8	8.6	2.2E-6	1.2E-3	
	cellular macromolecular complex assembly	8	8.6	2.7E-6	1.4E-3	
	regulation of cellular localization	7	7.5	3.5E-6	1.6E-3	
	regulation of actin filament polymerization	4	4.3	4.0E-6	1.8E-3	
	cellular macromolecular complex subunit organization	8	8.6	5.1E-6	2.5E-3	
	regulation of actin polymerization or depolymerization	4	4.3	5.6E-6	2.2E-3	
	regulation of actin filament length	4	4.3	6.1E-6	2.2E-3	
	regulation of synaptic plasticity	4	4.3	6.4E-6	2.2E-3	
	regulation of neurotransmitter secretion	3	3.2	6.7E-6	2.2E-3	
	vehicle-mediated transport	10	10.8	6.8E-6	2.2E-3	
	regulation of secretion	6	6.5	6.9E-6	2.2E-3	
	regulation of neurotransmitter levels	4	4.3	7.2E-6	2.2E-3	
	regulation of protein polymerization	4	4.3	7.6E-6	2.2E-3	
	synaptic transmission	7	7.5	8.4E-6	2.4E-3	
	regulation of neurotransmitter transport	3	3.2	1.0E-5	2.7E-3	
	positive regulation of transport	6	6.5	1.0E-5	2.7E-3	
	actin cytoskeleton organization	6	6.5	1.1E-5	2.7E-3	
	purine nucleoside biosynthetic process	5	5.4	1.1E-5	2.8E-3	
	negative regulation of actin filament polymerization	3	3.2	1.2E-5	2.8E-3	
	protein stabilization	3	3.2	1.2E-5	2.8E-3	
	nitrogen compound biosynthetic process	7	7.5	1.2E-5	2.8E-3	
	negative regulation of organelle organization	4	4.3	1.2E-5	2.8E-3	
	negative regulation of protein polymerization	3	3.2	1.2E-5	2.8E-3	
	nucleosome assembly	4	4.3	1.2E-5	2.9E-3	
	actin filament-based process	6	6.5	1.4E-5	2.9E-3	
	cytoskeleton organization	8	8.6	1.5E-5	2.9E-3	
	chromatin assembly	4	4.3	1.5E-5	2.9E-3	
	ATP biosynthetic process	4	4.3	1.6E-5	3.0E-3	
	regulation of actin-cytoskeleton organization	4	4.3	1.6E-5	3.0E-3	
	macromolecular complex assembly	10	10.8	1.6E-5	3.0E-3	
	protein-ORF complex assembly	4	4.3	1.7E-5	3.0E-3	
	regulation of exocytosis	3	3.2	1.7E-5	3.0E-3	
	negative regulation of protein complex assembly	3	3.2	1.7E-5	3.0E-3	
	regulation of actin filament-based process	4	4.3	1.7E-5	3.0E-3	
	transmission of nerve impulse	7	7.5	1.7E-5	3.0E-3	
	nucleosome organization	4	4.3	1.8E-5	3.0E-3	
	regulation of vesicle-mediated transport	4	4.3	1.9E-5	3.0E-3	
	sensory perception of sound	4	4.3	2.0E-5	3.0E-3	
	purine ribonucleoside triphosphate biosynthetic process	4	4.3	2.0E-5	3.0E-3	
	purine nucleoside triphosphate biosynthetic process	4	4.3	2.1E-5	3.0E-3	
	ribonucleoside triphosphate biosynthetic process	4	4.3	2.1E-5	3.0E-3	
	regulation of cellular component size	6	6.5	2.2E-5	3.0E-3	
	nucleoside triphosphate biosynthetic process	4	4.3	2.2E-5	3.0E-3	
	sensory perception of mechanical stimulus	4	4.3	2.2E-5	3.0E-3	
	positive regulation of neurotransmitter secretion	2	2.2	2.2E-5	3.0E-3	
	macromolecular complex subunit organization	10	10.8	2.4E-5	3.2E-3	
	ATP metabolic process	4	4.3	2.4E-5	3.2E-3	
	purine nucleoside metabolic process	5	5.4	2.4E-5	3.2E-3	
	nucleotide biosynthetic process	5	5.4	2.4E-5	3.2E-3	
	cell-cell signaling	9	9.7	2.5E-5	3.2E-3	
	regulation of protein stability	3	3.2	2.6E-5	3.2E-3	
	nucleobase, nucleoside, nucleotide and nucleic acid biosynthetic	5	5.4	2.7E-5	3.2E-3	
	nucleobase, nucleoside and nucleotide biosynthetic process	5	5.4	2.7E-5	3.2E-3	
	purine ribonucleoside biosynthetic process	4	4.3	3.2E-5	3.3E-3	
	ORF packaging	4	4.3	3.2E-5	3.3E-3	
	purine ribonucleoside triphosphate metabolic process	4	4.3	3.2E-5	3.3E-3	
	ribonucleoside triphosphate metabolic process	4	4.3	3.2E-5	3.3E-3	
	secretion by cell	5	5.4	3.4E-5	4.0E-3	
	positive regulation of neurotransmitter transport	2	2.2	3.5E-5	4.0E-3	
	purine nucleoside triphosphate metabolic process	4	4.3	3.6E-5	4.0E-3	
	posttranscriptional regulation of gene expression	5	5.4	3.6E-5	4.0E-3	
	ion transport	10	10.8	3.7E-5	4.5E-3	
	ribonucleoside biosynthetic process	4	4.3	3.7E-5	4.5E-3	
	chromatin assembly or disassembly	4	4.3	3.9E-5	4.2E-3	
	regulation of organelle organization	5	5.4	4.0E-5	4.2E-3	
	aspartate transport	2	2.2	4.1E-5	4.2E-3	
	negative regulation of cytoskeleton organization	3	3.2	4.2E-5	4.2E-3	
	nucleoside triphosphate metabolic process	4	4.3	4.2E-5	4.2E-3	
	cation transport	8	8.6	4.5E-5	4.4E-3	
	phosphorylation	10	10.8	4.6E-5	4.4E-3	
	regulation of cytoskeleton organization	4	4.3	4.7E-5	4.4E-3	
	purine ribonucleoside metabolic process	4	4.3	4.8E-5	4.5E-3	
	ribonucleoside metabolic process	4	4.3	5.4E-5	5.0E-3	
	neurotransmitter uptake	2	2.2	5.8E-5	5.0E-3	
	behavior	7	7.5	5.9E-5	5.1E-3	
	phosphorus metabolic process	11	11.8	6.0E-5	5.1E-3	
	phosphate metabolic process	11	11.8	6.0E-5	5.1E-3	
	regulation of binding	4	4.3	6.2E-5	5.2E-3	
	locomotory behavior	5	5.4	7.8E-5	6.0E-3	
	L-glutamate transport	2	2.2	8.0E-5	6.0E-3	
	dicarboxylic acid transport	2	2.2	8.5E-5	6.2E-3	
	RNA splicing	5	5.4	8.7E-5	6.3E-3	
	acidic amino acid transport	2	2.2	9.1E-5	6.4E-3	
	actin cytoskeleton reorganization	2	2.2	9.1E-5	6.4E-3	
	signal complex assembly	2	2.2	9.1E-5	6.4E-3	
	positive regulation of cellular component organization	4	4.3	9.2E-5	6.4E-3	
	intracellular transport	8	8.6	9.2E-5	6.4E-3	
	response to interleukin-1	2	2.2	9.6E-5	6.5E-3	
	regulation of cell projection organization	3	3.2	9.7E-5	6.5E-3	
	neurological system process	12	12.9	9.9E-5	6.5E-3	

Category	Term	Count	%	P-Value	Benjamini
GOTERM_MF_FAT	inorganic cation transmembrane transporter activity	11	11.8	2.8E-8	8.5E-6
	cytoskeletal protein binding	16	17.2	3.6E-7	5.3E-5
	monovalent inorganic cation transmembrane transporter activity	8	8.6	3.4E-6	3.2E-4
	hydrogen ion transmembrane transporter activity	7	7.5	1.9E-5	1.4E-3
	heme-copper terminal oxidase activity	5	5.4	2.4E-5	1.4E-3
	oxidoreductase activity, acting on heme group of donors, oxygen	5	5.4	2.4E-5	1.4E-3
	oxidoreductase activity, acting on heme group of donors	5	5.4	2.4E-5	1.4E-3
	cytochrome-c oxidase activity	5	5.4	2.4E-5	1.4E-3
	NADH dehydrogenase (quinone) activity	5	5.4	1.4E-6	6.7E-3
	NADH dehydrogenase activity	5	5.4	1.4E-6	6.7E-3
	NADH dehydrogenase (ubiquinone) activity	5	5.4	1.4E-6	6.7E-3
	actin binding	10	10.8	1.7E-6	7.2E-3
	oxidoreductase activity, acting on NADH or NADPH, quinone or a	5	5.4	2.2E-6	8.4E-3
	structural constituent of cytoskeleton	5	5.4	1.1E-6	3.6E-3
	calcium ion binding	15	16.1	1.2E-6	3.9E-3
	symporter activity	6	6.5	1.6E-6	4.2E-3
	L-amino acid transmembrane transporter activity	4	4.3	1.6E-6	3.9E-3
	oxidoreductase activity, acting on NADH or NADPH	5	5.4	1.7E-6	3.9E-3
	calmodulin binding	6	6.5	1.7E-6	3.9E-3
	L-glutamate transmembrane transporter activity	3	3.2	2.0E-6	3.8E-3
	acidic amino acid transmembrane transporter activity	3	3.2	2.4E-6	4.2E-3
	solute/cation symporter activity	5	5.4	2.8E-6	4.7E-3
	cytoskeletal adaptor activity	3	3.2	2.8E-6	4.5E-3
	amino acid transmembrane transporter activity	4	4.3	5.7E-6	8.5E-3
	SNAP receptor activity	3	3.2	7.2E-6	1.0E-1
	organic acid/sodium symporter activity	3	3.2	1.0E-5	1.2E-1
	amine transmembrane transporter activity	4	4.3	1.1E-5	1.2E-1
	di-, tri-valent inorganic cation transmembrane transporter activit	3	3.2	2.4E-5	2.7E-1
	P- β -bond-hydrolysis-driven transmembrane transporter activity	4	4.3	3.9E-5	3.9E-1
	primary active transmembrane transporter activity	4	4.3	3.9E-5	3.9E-1
	solute/sodium symporter activity	3	3.2	4.1E-5	3.9E-1
	enzyme binding	8	8.6	4.2E-5	3.8E-1
	channel regulator activity	3	3.2	5.1E-5	4.4E-1
	protein C-terminus binding	4	4.3	5.6E-5	4.6E-1
	lipid binding	7	7.5	5.9E-5	4.6E-1
	sodium/dicarboxylate symporter activity	2	2.2	6.0E-5	4.5E-1
	molecular adaptor activity	3	3.2	6.4E-5	4.7E-1
calcium ion transmembrane transporter activity	2	2.2	7.2E-5	4.9E-1	
ATPase activity, coupled to transmembrane movement of ions	3	3.2	7.6E-5	5.2E-1	
ATPase activity, coupled	5	5.4	8.7E-5	5.4E-1	
glucose transmembrane transporter activity	2	2.2	8.9E-5	5.4E-1	
sugar/hydrogen symporter activity	2	2.2	9.4E-5	5.5E-1	
cation/sugar symporter activity	2	2.2	9.4E-5	5.5E-1	
ion channel inhibitor activity	2	2.2	9.4E-5	5.5E-1	
dicarboxylic acid transmembrane transporter activity	2	2.2	9.4E-5	5.5E-1	
structural molecule activity	8	8.6	9.5E-5	5.5E-1	
channel inhibitor activity	2	2.2	1.0E-1	5.6E-1	
calcium channel regulator activity	2	2.2	1.0E-1	5.6E-1	